

ORIGINAL ARTICLE

Int22h-1/int22h-2-mediated Xq28 rearrangements: intellectual disability associated with duplications and in utero male lethality with deletions

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ABSTRACT

Background X linked intellectual disability (XLID) is common, with an estimated prevalence of 1/1000. The expanded use of array comparative genomic hybridisation (CGH) has led to the identification of several XLID-associated copy-number variants.

Methods Array CGH analysis was performed using chromosomal microarray with ~105 000 oligonucleotides covering the entire genome. Confirmatory fluorescence in situ hybridisation analyses were subsequently performed. Chromosome X-inactivation (XCI) was assessed using methylation-sensitive restriction enzyme digestion followed by PCR amplification.

Results A novel ~0.5 Mb duplication in Xq28 was identified in four cognitively impaired males who share behavioural abnormalities (hyperactivity and aggressiveness) and characteristic facial features (high forehead, upper eyelid fullness, broad nasal bridge and thick lower lip). These duplications were inherited from mothers with skewed XCI and are mediated by nonallelic homologous recombination between the low-copy repeat regions *int22h-1* and *int22h-2*, which, in addition to *int22h-3*, are also responsible for inversions disrupting the *factor VIII* gene in haemophilia A. In addition, we have identified a reciprocal deletion in a girl and her mother, both of whom exhibit normal cognition and completely skewed XCI. The mother also had two spontaneous abortions.

Conclusions The phenotypic similarities among subjects with *int22h-1/int22h-2*-mediated Xq28 duplications suggest that such duplications are responsible for a novel XLID syndrome. The reciprocal deletion may not be associated with a clinical phenotype in carrier females due to skewed XCI, but may be lethal for males in utero. Advancements in array CGH technology have enabled the identification of such small, clinically relevant copy-number variants.

linked intellectual disability (XLID), which has an estimated prevalence of 1/1000 males.^{1–4} To date, more than 200 different XLID disorders have been described, including around 150 syndromic forms, with over 80 XLID-associated genes having been identified (<http://xlmr.interfree.it>).

The expanded use of high-resolution genome analysis by array comparative genomic hybridisation (CGH) has led to the identification of several new microdeletion and microduplication syndromes.^{5–7} The application of arrays in XLID studies has resulted in the identification of new XLID-associated genes and copy-number variants (CNVs). For example, Froyen and colleagues screened a cohort of 108 subjects with ID by X-chromosome array CGH and identified X-chromosome CNVs in 14 subjects (13%).⁸ Duplications on the X chromosome correlated with ID more often than expected, suggesting a causal link between increased gene dosage and the disruption of normal cognitive development.³ The most common XLID-associated chromosomal aberrations are duplications of Xq28 comprising the *MECP2* gene, which have been reported in more than 100 cognitively impaired individuals with characteristic facial features, hypotonia, seizures, speech delay and recurrent infections.^{9–12} Other known X-chromosome CNVs associated with XLID include deletions of Xp11.23, Xp11.3, Xp11.4 and Xp22^{8 13–15} and duplications of Xp11.22, Xp11.22p11.23, Xp11.23, Xp11.4p21.3, Xp22.2 and Xq22.3.^{8 16–21}

Cytogenetically visible duplications at Xq are rare, with only around 40 reported cases.^{22 23} The most frequent are distal Xq duplications involving Xq26q28, which have been reported in patients with severe ID, hypotonia, genital malformation and characteristic facial features.^{24–27} Less common are interstitial duplications at Xq21q24 that have been reported in patients with developmental delay, severe feeding difficulties, growth retardation and digital anomalies.^{22 23} Most Xq duplications observed in males are inherited from a mother exhibiting a normal or near-normal phenotype. Less frequently, Xq duplications are found in females with ID.²³

We report four cognitively impaired males from three unrelated families with a novel ~0.5 Mb duplication in Xq28 located telomeric to the *MECP2* gene region and mediated by non-allelic homologous recombination between the low-copy

INTRODUCTION

The prevalence of intellectual disability (ID) in developed countries is estimated to be around 2%–3%, and approximately one-third of ID cases are thought to have a genetic aetiology. Of these genetic causes, about one-quarter may be attributed to mutations on the X chromosome resulting in X



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repeat (LCR) regions *int22h-1* and *int22h-2*, which are known to be involved in *factor VIII (F8)* gene inversions in subjects with severe haemophilia A. In addition, we report a reciprocal deletion in a mother and daughter with normal cognition. Herein, we describe the clinical phenotype of these subjects as well as the genomic structure of the deleted/duplicated region. In addition, we discuss the genes in this region and the potential effect of these rearrangements on the *F8* gene.

METHODS

Array CGH

Array CGH analysis was performed at the Medical Genetics Laboratories at Baylor College of Medicine. Informed consents were obtained as approved by the Institutional Review Board of Baylor College of Medicine. DNA was extracted from whole blood using the Puregene DNA extraction kit (Gentra, Minneapolis, Minnesota, USA) according to the manufacturer's instructions. The oligonucleotide-based chromosomal microarray version 7 (CMA V7 OLIGO) comprises approximately 105 000 oligonucleotides, which cover the entire genome at an average resolution of 30 kb with increased coverage at known disease loci. The array also includes six regions of known polymorphic variants.²⁸ The procedures for DNA digestion, labelling and hybridisation were performed according to the manufacturer's instructions with some modifications. The slides were scanned into image files using a GenePix Model 4000B microarray scanner (Molecular Devices, Sunnyvale, California, USA) or an Agilent G2565 laser scanner, after which the image files were quantitated using Agilent Feature extraction software (version 9.0). Text file outputs from the quantitation analysis were imported to an in-house analysis package for copy-number analysis.²⁹

Fluorescence in situ hybridisation analyses

Confirmatory fluorescence in situ hybridisation (FISH) analyses with bacterial artificial chromosome clones were performed on peripheral blood lymphocytes using standard procedures following the detection of copy-number changes via array CGH.³⁰

Chromosome X-inactivation studies

Chromosome X-inactivation (XCI) studies were performed at the *AR* (androgen receptor)³¹ and *FMR1* (fragile X mental retardation 1) loci.³² One hundred nanograms of genomic DNA from peripheral leucocytes of each female were digested with and without the methylation-sensitive restriction enzyme *HpaII* (New England Biolabs). DNA samples from male individuals were analysed without *HpaII* digestion. Five nanograms of DNA from each sample were subjected to PCR amplification with primers flanking the *AR* (CGA)_n repeat region (5'-AC-CAGGTAGCCTGTGGGGCCTCTACGATGGGC3' forward and 5'-CCAGAGCGTGCGGAAGTGATCCAGAACCCGG3' reverse) and the *FMR1* (CGG)_n repeat region as previously described.^{31 32} PCR products were separated on an ABI 3770 Analyser and analysed with GeneMapper software. The XCI ratio was calculated as previously described.³³ Inactivation ratios greater than 80:20 were designated as skewed XCI, whereas ratios greater than 95:5 were considered extremely skewed XCI.

Bioinformatics and in silico sequence analysis

Genomic sequence data based on the oligonucleotide coordinates from array CGH analysis were downloaded from the UCSC genome browser (Build 36, UCSC genome browser, March 2006). Regional assemblies were aligned using NCBI BLAST (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>) in order to identify the LCR regions.

RESULTS

Clinical description

Family 1 includes two brothers and their mother. The older brother is an 11-year-old who was born at term with a birth weight of 3.7 kg (50th–75th percentile) and a length of 56 cm (just above the 97th percentile). The pregnancy was complicated by gestational diabetes and cigarette smoking. During infancy, he experienced three episodes of pneumonia, all of which required hospitalisations. He was noted to be developmentally delayed beginning in early childhood. He started talking at the age of 6 years, started walking at 3 years and was toilet trained at 7 years. He is enrolled in special education classes and undergoes physical, occupational and speech therapy. At the time of this report, he was able to recognise letters and numbers and could write his name. However, he cannot read or solve simple mathematical problems. He exhibits autistic behaviours, including minimal eye contact and rocking, and was diagnosed as having Asperger syndrome. He also exhibits abnormally aggressive behaviours, including self-biting and hitting others. In addition, he presented with toe-walking, which required heel-cord lengthening, recurrent nosebleeds requiring multiple cauterisations, recurrent ear infections requiring ear tube placement, reactive airway disease and difficulty sleeping. Previous evaluations, including brain MRI, fragile-X DNA testing, and urine amino and organic acids, were all normal. Physical examination revealed a weight and height 3 SD above the mean and a head circumference at the 90th percentile. Distinctive facial features were also noted (table 1, figure 1A).

His younger brother is a 3-year-old born at term with a birth weight and length at about the 50th percentile. This pregnancy was also complicated by gestational diabetes and cigarette smoking. During the neonatal period, he was found to have a murmur, and an echocardiogram showed patent foramen ovale and patent ductus arteriosus, both of which had closed spontaneously prior to a subsequent echocardiogram. Similar to his older brother, he exhibited delays in attaining his developmental milestones. He started walking at 18 months of age and talking at 2 years. At the age of 3 years, he is only able to talk in simple two-word sentences. He receives physical and speech therapy. In addition, he has bilateral metatarsus adductus, requiring braces, and recurrent ear infections, which required ear tube placement. He also failed the newborn hearing screen and two subsequent hearing tests, and is being evaluated for a hearing aid. Physical examination revealed normal growth parameters and some distinctive facial features (table 1, figure 1B).

They have two maternal half-sisters, a 5-year-old enrolled in special education and a 14-year-old honour student, as well as two healthy 3- and 5-year-old paternal half-brothers. The parents are non-consanguineous. The mother is 32 years old and was reported to have learning difficulties. She required special education and did not continue schooling beyond the 11th grade. She was noted to have some similar facial features to her sons (table 1, figure 1C). The father has been healthy and did not report learning difficulties. The family history also revealed maternal uncles who required special education (figure 2).

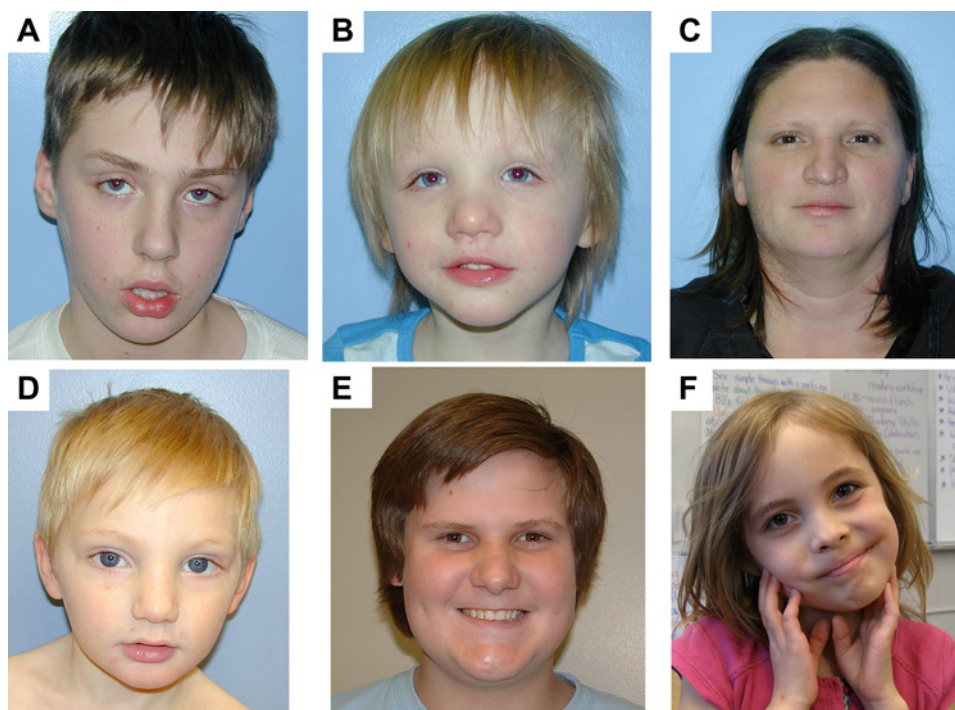
Family 2 includes a 3-year-old boy evaluated for developmental delay and microcephaly, and his mother. He was born to a 16-year-old mother who reported usage of birth control pills for the first few months of pregnancy and had premature contractions at 6 months of pregnancy, which required hospitalisation. He was born at term with a birth weight around the 50th percentile and stayed in the neonatal intensive care unit because of hypoglycemia and jaundice. The mother had

Table 1 Clinical features, factor VIII levels and chromosome X-inactivation results for subjects with *int22h-1/int22h-2*-mediated Xq28 duplications and deletions

Chromosome X-inactivation											
					AR locus		FMR1 locus				
					FVIII (%)	Allele size (nt)	Ratio	Allele size (nt)	Ratio		
ID	Facial features				Behavioural abnormalities	Other clinical manifestations					
Xq28 duplication Family 1	Older brother 11 years	+	High forehead, long face, upper eyelid fullness, open mouth, thick lower lip, narrow high arch palate, teeth crowding and overbite (figure 1A).		Asperger syndrome, aggressive behaviours and difficulty sleeping.	Recurrent pneumonias and ear infections, nosebleed, reactive airway disease and toe-walking.	124	248	NA	281	NA
	Younger brother 3 years	+	High forehead, upper eyelid fullness, broad nasal bridge, sparse eyebrows, small ears with simple helices, thick lower lip and tongue tie with lingual frenulum (figure 1B)		—	Bilateral metatarsus adductus, recurrent ear infections, hearing loss, and patent foramen ovale and ductus arteriosus.	286	248	NA	380	NA
	Mother 32 years	+	High forehead, broad nasal bridge, sparse eyebrows and thick lower lip (figure 1C).		—	—	NA	239 /248	89:11	281/380	13:87
Family 2	Son 3 years	+	High forehead, upper eyelid fullness, deep-seated eyes, broad nasal bridge, thick lower lip and retrognathia (figure 1D).		Hyperactivity and aggression.	Pneumonia and recurrent ear infections.	268	242	NA	278	NA
	Mother 19 years	+	NA		NA	NA	NA	242/254	12:88	278/308	13:87
Family 3	Son 15 years	+	High forehead, upper eyelid fullness, deep-seated eyes, broad nasal bridge and retrognathia (figure 1E).		Motor tics, anxiety and attention deficit hyperactivity disorder.	Recurrent ear infections, asthma, allergic rhinitis, joint pains, mild proximal muscle weakness, hypotonia, flat feet, recurrent nosebleed, easy bruising and prolonged bleeding.	24	242	NA	314	NA
	Mother 41 years	+	NA		Depression.	Hypothyroidism.	NA	242/242	NI	308/314	6:94
Xq28 deletion Family 4	Daughter 6 years	—	High forehead (figure 1F).		Hyperactivity, inattentive, impulsiveness, stereotypic movements and sensory integration difficulties.	Clinodactyly and café-au-lait spot.	60	254/257	0:100	Not done	NA
	Mother 39 years	—	NA		—	NA	NA	233/257	0:100	Not done	NA

The preferentially inactive alleles in females are in bold.
 Chromosome X-inactivation assays were performed at the *AR* (androgen receptor) and *FMR1* (fragile X mental retardation 1) loci.
 ID, intellectual disability or developmental delay; FVIII, factor VIII levels with normal range 50%–200%; NA, not available or applicable; NI, non-informative; nt, nucleotides.

Figure 1 Facial features in individuals with *int22h-1/int22h-2*-mediated Xq28 rearrangements. (A) Facial features in the older brother in family 1, including high forehead, long face, upper eyelid fullness, open mouth and thick lower lip. Narrow, high arched palate, teeth crowding and overbite are not shown. (B) Facial features in the younger brother in family 1, including high forehead, upper eyelid fullness, broad nasal bridge, sparse eyebrows and thick lower lip. Small ears with simple helices and tongue tie with lingual frenulum are not shown. (C) Facial features in the mother in family 1, including high forehead, broad nasal bridge, sparse eyebrows and thick lower lip. (D) Facial features in the proband in family 2, including high forehead, upper eyelid fullness, deep-seated eyes, broad nasal bridge, thick lower lip and retrognathia. (E) Facial features in the proband in family 3, including high forehead, upper eyelid fullness, deep-seated eyes, broad nasal bridge and retrognathia. (F) Facial features in the proband in family 4, including high forehead, with no other dysmorphic features.



difficulty bonding with the baby, and he was therefore discharged with a foster family and was subsequently adopted. During the first few months of life, he was noted to have microcephaly. A skull x-ray was performed at the age of 9 months and showed no evidence of craniosynostosis. His medical history is significant for recurrent ear infections and a hospitalisation at the age of 7 months for pneumonia. Developmentally, he showed mild global delay, including rolling over at the age of 8 months, followed by cooing and sitting without support at 9 months. An assessment at 12 months indicated a developmental level of 9 months. Therefore, physical and speech therapy was initiated. By the age of 3 years, hyperactivity and aggression were noted, and physical examination revealed a head circumference within the third–fifth percentiles with height and weight around the 50th percentiles. He was also noted to have some distinctive facial features (table 1, figure 1D). He has a healthy 4-year-old maternal half-brother. No medical history was available for the biological father. His mother, who was reported to have learning difficulties, has a full brother and maternal half-brother, both of whom are intellectually disabled (figure 2).

Family 3 includes a boy and his mother. This 15-year-old male was born at term weighing 4.7 kg with a length of 55.9 cm, both of which are above the 95th percentiles. He was noted to be developmentally delayed during early childhood and was subsequently enrolled in special education as well as physical, occupational and speech therapy. Cognitive assessment at the age of 14 years revealed low ability across all cognitive domains with a full-scale IQ of 57. In addition, he has motor tics manifesting as head and neck twitching, has anxiety and was diagnosed as having attention deficit hyperactivity disorder. His medical history is significant for asthma, allergic rhinitis and recurrent ear infections requiring tube placement and adenotonsillectomy at the age of 5 years. For the past several years, he has been having recurrent episodes of erythema and oedema in

his distal upper and lower extremities, joint pains, headaches and progressive muscle weakness in the forms of decreased exercise capacity and general slowness. Brain and spinal MRIs were normal, and a rheumatologic evaluation consisting of anti-phospholipid and anticardiolipin antibodies was negative. He also presented with recurrent nosebleeds requiring repeated cauterisation, easy bruising and delayed clotting following skin laceration. His weight is 84 kg (>95th percentile), his height is 172.7 cm (75th–90th percentiles), and his head circumference is 56.7 cm (75th–90th percentiles). Additional findings on physical examination include lower extremity oedema, mild proximal muscle weakness and hypotonia with normal muscle bulk and reflexes, flat feet and distinctive facial features (table 1, figure 1E). As a result of his bleeding tendency, the factor VIII level was tested and found to be 24%, which is consistent with mild haemophilia A (table 1). A subsequent factor VIII inhibitor assay was negative, whereas VWF antigen and activity were within normal ranges. As part of factor VIII deficiency evaluation, *F8* gene inversion analysis was performed using the method described by Liu and colleagues³⁴ and was reported as positive for an *F8* inversion. However, *F8* gene inversions are associated with severe haemophilia A, which is inconsistent with the mild phenotype observed in this patient. Consequently, we realised that these results were misinterpreted for this patient as a result of the inherent limitations of *F8* gene inversion analysis in the presence of an Xq28 duplication. This inversion analysis is performed by multiple PCR using four primers to differentiate between the normal *F8* locus, inversions and carrier females (figure 3A–C). Although this patient does not harbour an *F8* inversion, the results were misinterpreted as positive for an *F8* inversion due to the presence of the Xq28 duplication (figure 3D). Subsequent sequencing of the *F8* gene revealed hemizygosity for the point mutation c.6089G>A (p.S2030N), which has been associated with mild haemophilia A (<http://hdb.org.uk/>).³⁵ The identification of this mutation explains the patient's mild

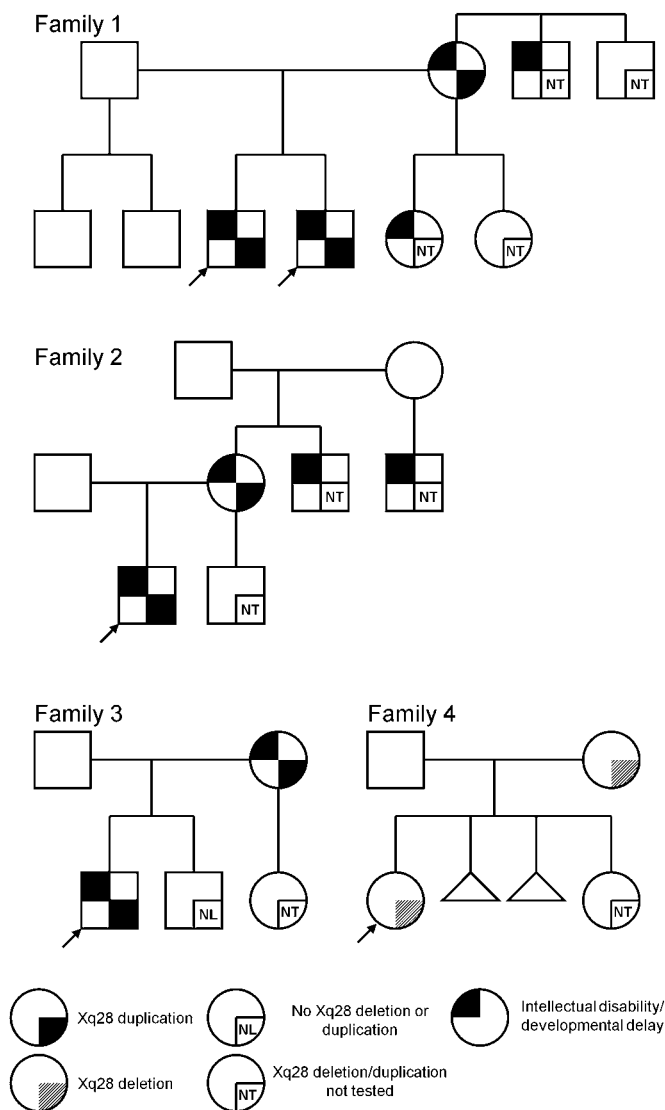


Figure 2 Pedigrees for the three reported families with Xq28 duplications and the fourth family with the reciprocal Xq28 deletion.

haemophilic phenotype. An additional investigation included a karyotype, fragile X DNA testing, CPK level assay and thyroid function tests, all of which were normal. He has a healthy brother and maternal half-sister. His 41-year-old mother was reported to have learning difficulties, obesity, depression and hypothyroidism. The father is 42 years old and has hypothyroidism (figure 2).

Family 4 includes a girl and her mother. The 6-year-old girl was born at term after an uncomplicated pregnancy. She gained her developmental milestones appropriately during early childhood, but concerns have arisen regarding certain behaviours, including hyperactivity, inattentiveness, impulsivity, stereotypic movements (eg, running in circles and arm-flapping) and sensory-seeking behaviours (eg, licking her fist and putting her fingers in her mouth). Neurodevelopmental evaluation at the age of 5 years revealed sensory integration difficulties, for which occupational therapy was recommended. Beginning at 3 years old, she was also observed to have staring episodes. Brain MRI and continuous EEG monitoring were normal, and it was concluded that these episodes are non-epileptic in nature and are likely due to inattention. Physical examination revealed normal growth parameters, clinodactyly, a café-au-lait spot on her back and a high forehead with no dysmorphic features (table 1, figure 1F).

Previous investigation, including fragile X DNA testing and thyroid function tests, was normal. She has a 1-year-old healthy sister. Her mother is a 39-year-old who has been healthy with normal intelligence, but had two spontaneous abortions at 5 and 8 weeks' gestation. Her father is 44 years old with depression and elevated cholesterol (figure 2).

Array CGH and FISH analyses

CMA V7 OLIGO performed on the four probands of families 1–3 revealed ~0.5 Mb Xq28 duplications (153.7–154.2 Mb, hg18), which were confirmed by FISH analysis in each proband. The mothers in families 1–3 were also found to carry the same duplications by FISH. Furthermore, the proband's brother in family 3 was found to have a normal CMA. The proband in family 4 was found to carry a reciprocal Xq28 deletion and an ~1 Mb 15q25.3 duplication (83.4–84.4 Mb, hg18). FISH determined that her father carries the 15q25.3 duplication, while her mother carries the Xq28 deletion (figures 2 and 4).

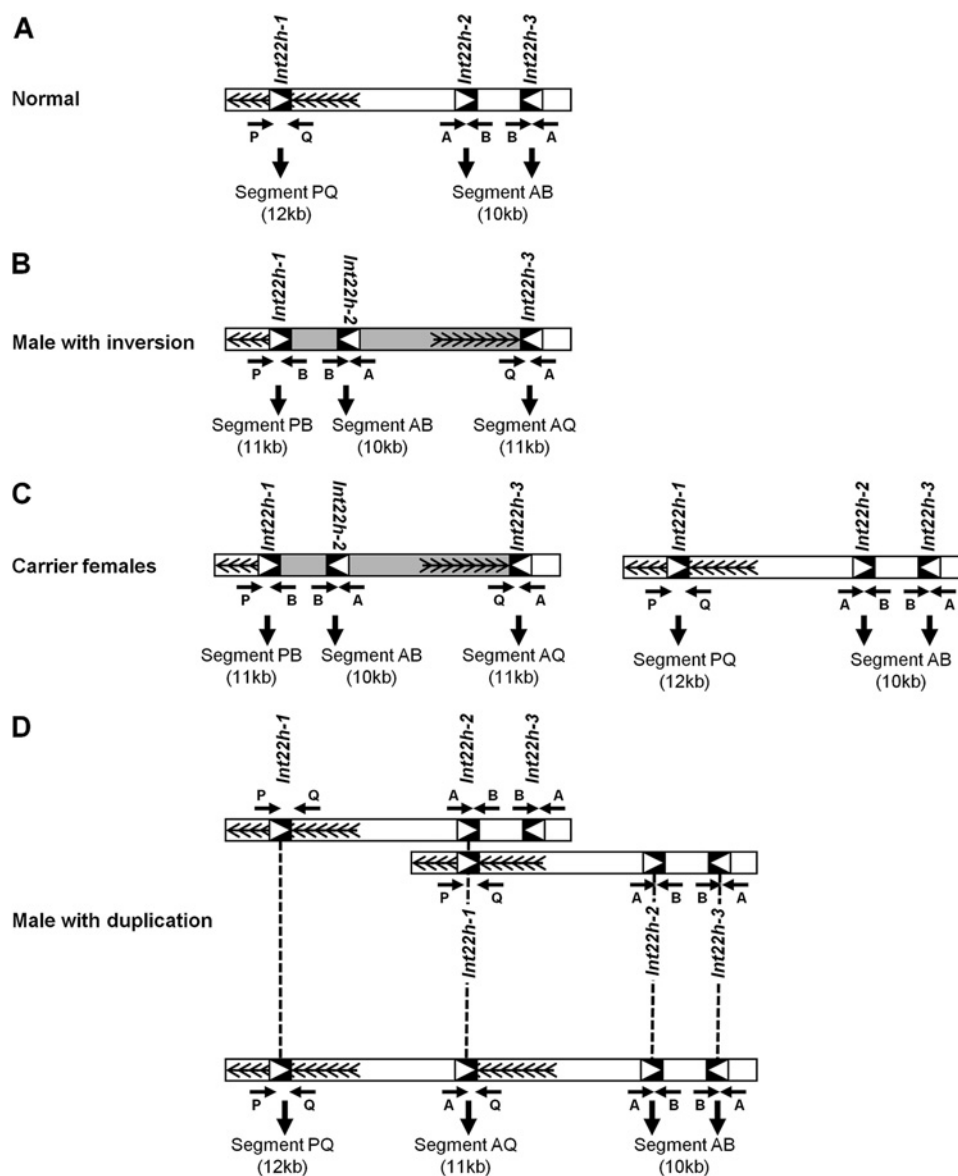
XCI assay

XCI was assessed in the mothers of families 1, 2 and 3 at the *AR* and *FMR1* loci whereby skewed XCI patterns were observed in all three mothers (table 1). Interestingly, the brothers in family 1 inherited the same *AR* allele (248) but opposite *FMR1* alleles (281 vs 380). The *AR* 248 and the *FMR1* 281 alleles represent the preferentially active X chromosome in their mother, and it appears that both sons did inherit this same homologue but with one of them carrying the *FMR1* 380 allele most likely due to a recombination event. The finding that the preferentially active chromosome in the mother of family 1 is the same allele carried by her son (*AR* 242 and *FMR1* 278), which is consistent with the findings in family 1. That is, the normal X chromosome is preferentially inactivated, while the preferentially active X chromosome is that which carries the Xq28 duplication. Although the XCI assay at the *AR* locus was not informative for the mother of family 3, the assay at the *FMR1* locus indicated preferential inactivation of the same allele carried by her son, suggesting that the X chromosome carrying the Xq28 duplication is preferentially inactivated in the mother. The daughter and mother in family 4 carrying the reciprocal Xq28 deletion showed 100% skewing XCI at the *AR* locus (table 1).

Xq28 genomic architecture

Using Blast2, we analysed the DNA sequence in the Xq28 region between 153.7 and 154.4 Mb, which harbours the duplication/deletion breakpoints, searching for alignments at least 2 kb in length with greater than 90% DNA sequence identity. The region contains three ~9.5 kb LCR copies with 99.7% DNA sequence identity, one in intron 22 of the *F8* gene and two located distally, which are consistent with the previously described *int22h-1*, *int22h-2* and *int22h-3* regions.³⁶ The deletion and duplication breakpoints were localised within the directly oriented *int22h-1* and *int22h-2* regions. We identified two ~50 kb LCR copies with 98.7% of the DNA sequence consistent with the previously described palindrome arms.³⁶ In addition to these previously described regions, we also identified 10 LCR copies ~5–6 kb in size and of 90%–94% DNA sequence identity (figure 5).

Figure 3 Schematic representation of the Xq28 region demonstrating the *F8* gene using arrowheads and *int22h-1*, 2 and 3 as black boxes with white triangles indicating their relative orientations.^{36–40} Panel (A) represents the normal genome, whereas panels (B–D) represent the results of different recombination events between *int22h-1*, 2 and 3, and their subsequent effects on the *F8* inversion molecular testing described by Liu and colleagues.³⁴ Two primers, P and Q, are located within the *F8* gene at positions -1212 base pairs (bp) and +1334 bp flanking *int22h1*. Two primers, A and B, are located at -167 bp and +118 bp flanking *int22h2* and *int22h3*. (A) In a normal male, segments PQ (12 kb) and AB (10 kb) are produced. (B) The *int22h2/int22h3*-mediated inversion in males with haemophilia results in production of PB (11 kb) and QA segments (11 kb), along with the AB segment (10 kb) from the non-recombined *int22h-2*. (C) Females heterozygous for the inversion (carriers) produce PB (11 kb), QA (11 kb) and AB (10 kb) segments from the inverted allele, and PQ (12 kb) and AB (10 kb) from the normal allele. (D) Diagrammatic demonstration of non-allelic homologous recombination between *int22h-1* and *int22h-2*, resulting in an *int22h-1/int22h-2*-mediated Xq28 duplication, which will result in formation of the AQ segment (11 kb) in addition to PQ (12 kb) and AB (10 kb), resulting in a pattern similar to that of a female heterozygous for the inversion.



Factor VIII assessments

Factor VIII levels were assayed in all probands. Only the proband in family 3 showed a reduced factor VIII level consistent with mild haemophilia A (table 1).

DISCUSSION

In this report, we present the first evidence for the presence of *int22h-1/int22h-2*-mediated Xq28 rearrangements with an ~0.5 Mb duplication in four cognitively impaired males who share behavioural abnormalities and characteristic facial features. The reciprocal deletion was detected in a cognitively normal girl and her mother.

The described rearrangement breakpoints were localised to the directly oriented 9.5 kb LCRs *int22h-1* and *int22h-2*. The *int22h-1* maps to intron 22 of the *F8* gene, while *int22h-2* and *int22h-3* are located distally. Genomic inversions between *int22h-1* and either *int22h-2* or *int22h-3* disrupt the *F8* gene in nearly half of severe haemophilia A cases such that it was initially assumed that *int22h-2* and *int22h-3* were in opposite orientation to *int22h-1*.^{37–38} However, the publication of the sequence of the human X chromosome in 2005 revealed that *int22h-1* and *int22h-2* are, in fact, oriented in the same direction and are in opposite orienta-

tion to *int22h-3*.³⁹ These same data also revealed that *int22h-2* and *int22h-3* are part of a palindrome, suggesting that recombination between the arms of this palindrome may establish a non-deleterious inversion polymorphism, which changes the relative positions and orientations of *int22h-2* and *int22h-3*, thus permitting inversions involving *int22h-1* and *int22h-2*. The low frequency of this polymorphic allele may explain the rarity of such inversions.^{40–41} Knowing that *int22h-1* and *int22h-2* are directly oriented, it has been predicted that recombination between *int22h-1* and *int22h-2* may also result in deletions and duplications. However, because of a lack of evidence, it has been suggested that they either are extremely rare or do not exist.^{36–39–42} Herein, due to advancements in high-resolution array CGH technology, we present evidence of such rearrangements in three families with *int22h-1/int22h-2*-mediated Xq28 duplication and in one family with the reciprocal deletion.

The *int22h-1/int22h-2*-mediated Xq28 duplications are approximately 0.5 Mb in size and are located telomeric to the *MECP2* gene region. All four boys (families 1–3) with this duplication share characteristics that include cognitive impairment, behavioural problems (hyperactivity and aggressiveness), recurrent infections (pneumonia and ear infections) and

characteristic facial features including high forehead, upper eyelid fullness, broad nasal bridge and thick lower lip (table 1, figure 1), suggesting that these duplications result in a distinct XLID syndrome. Furthermore, the absence of this duplication in the cognitively normal brother of the proband in family 3 lends additional support to this proposal. While the proband in family 3 also exhibits relative microcephaly, it is unclear whether the duplication is responsible.

The duplications in the probands in families 1–3 were all maternally inherited, and all three mothers were reported to have learning difficulties. Additionally, the mother of family 1 shared some of the facial characteristics present in the four male probands, including high forehead, broad nasal bridge and thick lower lip (table 1, figure 1). These observations suggest that *int22h-1/int22h-2*-mediated Xq28 duplications can be associated with intellectual impairment in carrier females.

XCI assessments are typically performed using primers at the *AR* locus (Xq12).³¹ This method can very accurately determine whether XCI is skewed or random. However, this method has limited utility in predicting whether the preferentially active or inactive X chromosome harbours the Xq28 duplication since the large distance between the *AR* locus and Xq28 (about 87 Mb) suggests a high likelihood of recombination. Therefore, we have also performed the assay at the *FMR1* locus (Xq27), which is approximately 0.8 Mb from the duplicated region with a recombination frequency of less than 1%. Even so, the brothers in family 1 were found to have opposite alleles when they were tested at the *FMR1* locus, suggesting a recombination event. We used both assays to predict the XCI status and to determine whether the X chromosome carrying the duplication preferentially remains active or is inactivated. Ultimately, the mothers in families 1–3 all demonstrated skewed XCI, and the results were concordant between the two methods (table 1). The XCI results also suggest that the normal X chromosomes are preferentially inactivated in the mothers of families 1 and 2. Most females with duplications involving the X chromosome demonstrate skewed XCI whereby the abnormal X chromosome is preferentially inactivated, which explains why the majority of carrier females do not manifest clinically. However, skewed XCI in favour of the X chromosome harbouring the duplication has been reported in females with Xp11.22p11.23 duplications who also exhibit a clinical phenotype attributed to the duplication.¹⁸ Although the mechanisms for such inactivation patterns are currently unknown, increased expression of a gene or genes in the duplicated region may provide unknown advantages to dividing cells.¹⁸ Thus, *int22h-1/int22h-2*-mediated Xq28 duplication may act in a similar manner to the Xp11.22p11.23 duplications such that the X chromosome carrying the duplication remains preferentially active. The fact that this abnormal X chromosome remains preferentially active in the mothers of families 1 and 2 may provide an explanation for the phenotypic effect of such duplications on those carrier females. However, the X chromosome carrying the duplication is found to be preferentially inactivated in the mother of family 3 who was also reported to have learning difficulties.

It is unclear why the X chromosomes that carry the duplication remain preferentially active in the mothers of families 1 and 2, but not in the mother of family 3. Similarly, it is also unknown why the mother of family 3 exhibits learning difficulties despite the abnormal X chromosome being preferentially inactivated. One possible explanation is a recombination event between the *FMR1* locus and the Xq28 duplication region. The transposition of the duplication region to the opposite homologue would, in fact, yield results contrary to those of the XCI

assessment performed at the *FMR1* locus. This is because the duplication in the carrier mother would now be associated with *FMR1* 308 allele, which represents the active maternal chromosome. An alternative explanation involves the use of peripheral leucocytes for the XCI assays, such that the results exclusively reflect the XCI status of haematopoietic cells. It is possible that a distinct pattern of XCI occurs in the nervous system where the duplicated X chromosome may remain preferentially active. Such tissue-specific variation of XCI may provide a feasible explanation of the learning difficulties observed in the mother of family 3.^{33–42} Whereas our data are more consistent with the interpretation that the X chromosome carrying the Xq28 duplication remains preferentially active in the mothers of families 1 and 2, the observation of learning difficulties in the mother of family 3, whose Xq28 duplication is located on the inactive X chromosome, would argue against the significance of such genotype–phenotype correlation. Therefore, given the current inability to confirm any of the above theories, one cannot conclude decisively that the mothers' phenotypes are directly associated with an active abnormal X chromosome. Further studies of females with these duplications including assessment of X-inactivation status using additional markers within, and distal to, the duplication are needed to draw accurate conclusions as to the effect of such a duplication on the clinical phenotype and XCI in carrier females.

The mother and daughter in family 4 carry an *int22h-1/int22h-2*-mediated Xq28 deletion, although both are reported to have normal intelligence with no significant dysmorphic facial features. Since XCI showed a 100% skewed pattern in both individuals, these data suggest that the abnormal X chromosome is completely inactivated, such that these deletions are likely to have no phenotypic effect in females. However, it was noted that the mother had two spontaneous abortions. This observation coupled with the fact that this deletion has not been identified in males suggests that such deletions may be lethal for males in utero. Although family 4 includes the first cases of an *int22h-1/int22h-2*-mediated Xq28 deletion, one previous report indicated the presence of this deletion in a multi-generation family in which carrier females exhibited skewed XCI and experienced a higher than average rate of spontaneous abortion. The deletions were detected by FISH, and the breakpoints were roughly mapped using Southern analysis.^{43–44} The significant correlation between Xq28 deletion carrier status and the number of spontaneous abortions may support our hypothesis that this deletion is lethal for males in utero. Traditionally, chromosomal analysis has been recommended for females with recurrent spontaneous abortions, but *int22h-1/int22h-2*-mediated Xq28 deletions are beyond the resolution of standard karyotyping. We argue that these deletions can lead to recurrent spontaneous abortions in carrier females and therefore suggest considering array CGH as part of the investigation for multiple spontaneous abortions.

It has also been suggested that females carrying *int22h-1/int22h-2*-mediated Xq28 deletions should not exhibit clinical signs of haemophilia A as a result of the preferential inactivation of the X chromosome harbouring the *F8* gene-inclusive deletion. In addition, it was proposed that males or females with *int22h-1/int22h-2*-mediated Xq28 duplications would not be at risk of haemophilia A because they maintain a normal copy of the *F8* gene.⁴¹ Due to a lack of such cases, however, these hypotheses have not been tested until now. The patients reported here support the abovementioned suggestions. The daughter and mother in family 4 with the *int22h-1/int22h-2*-mediated Xq28

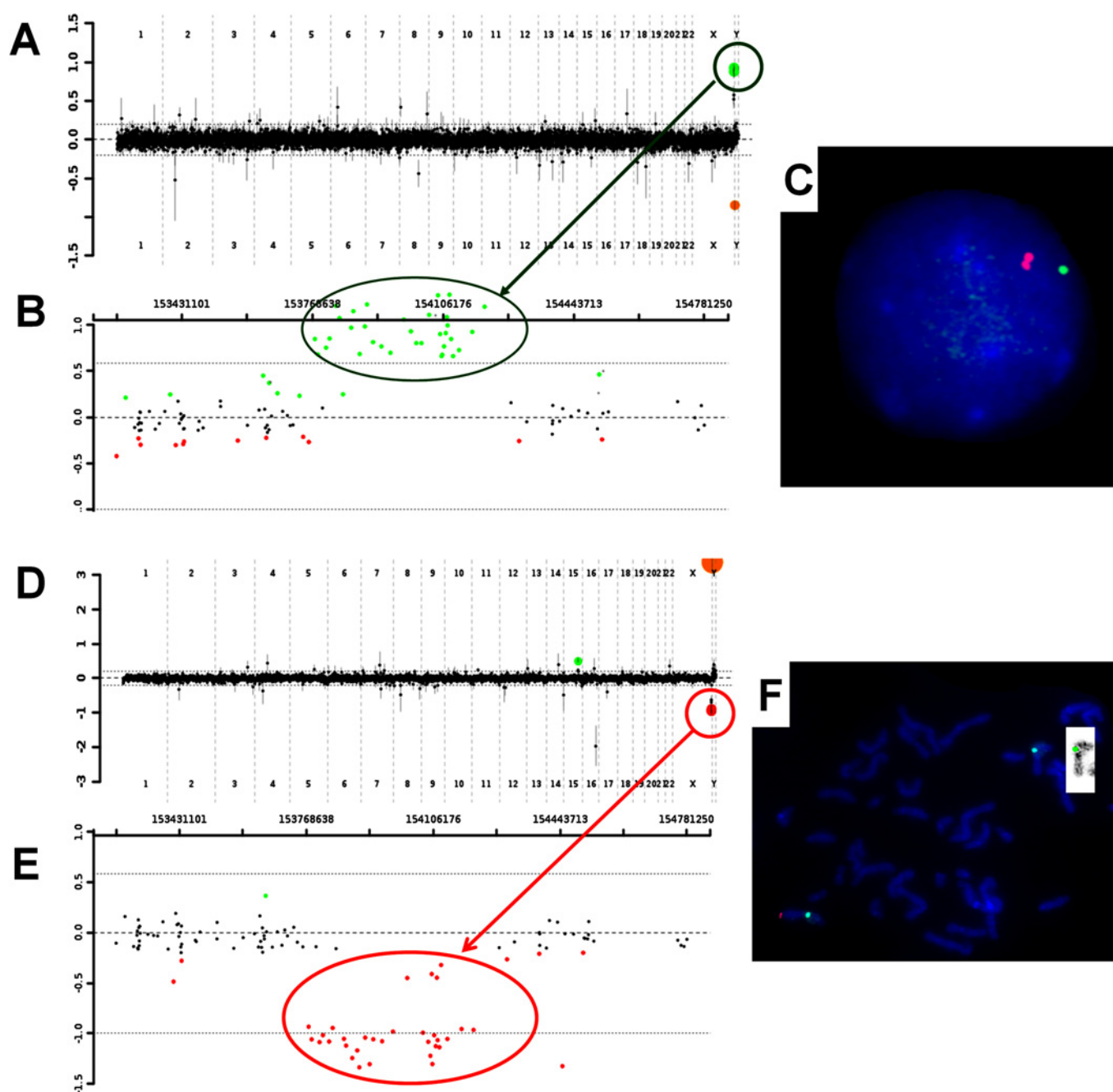


Figure 4 Panels (A), (B) and (C) demonstrate the chromosomal microarray and fluorescence in situ hybridisation (FISH) results for the proband of family 2. (A) Oligonucleotides are represented as green dots within the circle and indicate a gain of chromosome Xq28 material. (B) The Xq28 region, displaying individual oligonucleotides (green dots) that are displaced upward in the duplicated region. The proximal breakpoint is located between 153,759,353–153,774,852, and the distal breakpoint is located between 154,213,569–154,281,857. (C) FISH analyses showing the two bacterial artificial chromosome (BAC) clones, RP11-143H17 located within the duplicated region (red) and RP11-137H15 located proximal to the duplicated region and serving as a control (green). Panels (D), (E) and (F) demonstrate the chromosomal microarray and FISH results for the proband of family 3. (D) Oligonucleotides are represented as red dots within the circle and indicate a loss of chromosome Xq28 material. (E) The Xq28 region, displaying individual oligonucleotides (red dots) that are displaced downward in the deleted region. The proximal breakpoint is located between 153,759,353–153,774,852, and the distal breakpoint is located between 154,213,569–154,281,857. (F) FISH analyses showing the BAC clone RP11-143H17 (red) and the control probe targeting the centromere region of the X chromosome (green). The insert represents the reversed FISH image showing the deleted X chromosome.

deletion did not report any bleeding tendency, and the daughter was found to have a normal factor VIII level. In addition, the probands in families 1 and 2 who harbour the reciprocal duplication did not present with significant bleeding tendencies or low factor VIII levels.

Interestingly, the proband in family 3 does demonstrate a bleeding tendency and was found to have a low factor VIII

level consistent with mild haemophilia A. However, as previously indicated, the duplication should not result in a lower factor VIII level. Moreover, recurrent *F8* gene inversions are generally associated with severe haemophilia A, which conflicts with the mild phenotype observed in this patient. It has been proposed that *int22h-1/int22h-2*-mediated Xq28 duplications may confound the molecular diagnosis of haemophilia A.⁴⁵

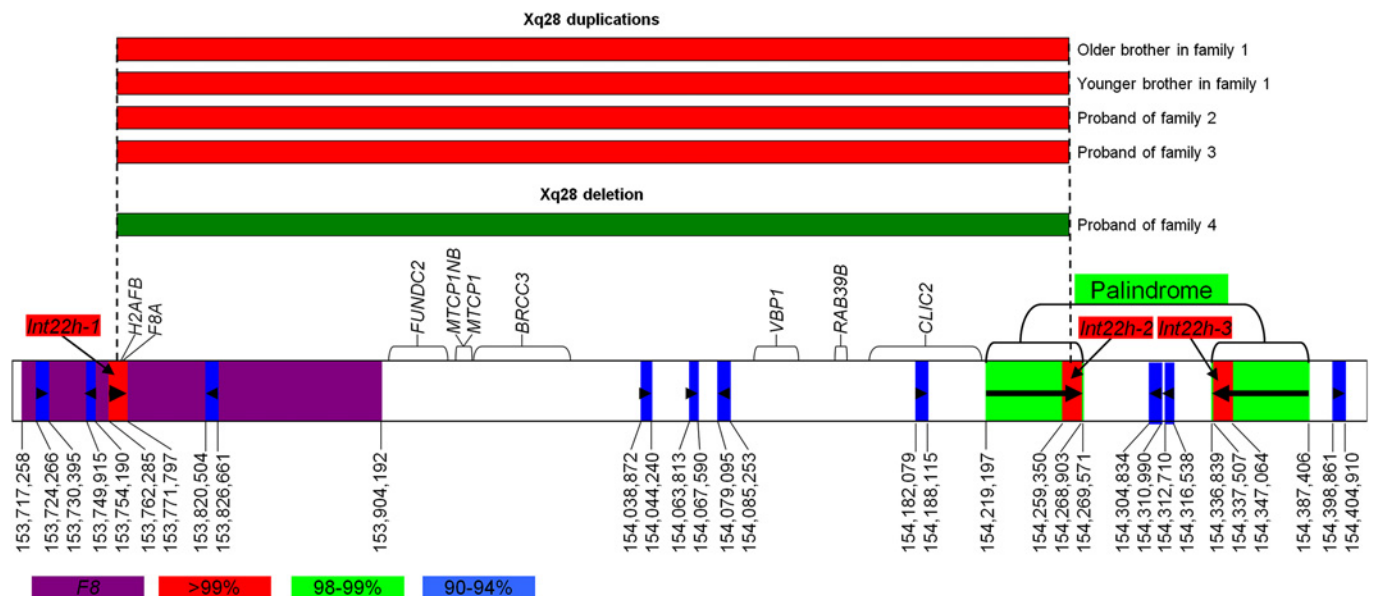


Figure 5 Schematic representation of the Xq28 region (153.7 and 154.4 Mb). The ~0.5 Mb Xq28 duplicated/deleted region with minimum size of 0.439 Mb (153,774,852–154,213,569) and maximum size of 0.523 Mb (153,759,353–154,281,857) is represented as a red rectangle for each duplication and a green rectangle for the deletion. The *F8* gene (153,717,258–153,904,192) is represented as a purple rectangle. The low-copy repeat (LCR) regions with DNA sequence identity >99% are also represented as red rectangles, the LCR regions with a DNA sequence identity of 98%–99% are represented as green rectangles, and the LCR regions with a DNA sequence identity of 90%–94% are represented as blue rectangles. The *int22h-1* region is 9512 base pairs (bp) and spans 153,762,285–153,771,797; the *int22h-2* region is 9553 bp in length and spans 154,259,350–154,268,903, and the *int22h-3* region is 9557 bp in length and spans 154,337,507–154,347,064. The proximal palindrome arm is 50374 bp in length and spans 154,219,197–154,269,571, and the distal palindrome arm is 50567 bp in length and spans 154,336,839–154,387,406.

Indeed, this was found to be true in the proband in family 3, whose *F8* gene inversion analysis was performed using multiple PCR assay to identify inversions responsible for haemophilia A.³⁴ Using this method, *int22h-1/int22h-2*-mediated Xq28 duplications result in a pattern indicative of a female inversion carrier (figure 3D). Therefore, this patient's results were falsely interpreted as positive for an *F8* inversion. Considering that these duplications should not affect *F8* gene expression, as well as the inherent limitations of *F8* gene inversion analysis in the presence of such duplications, *F8* gene sequencing was performed as a means of explaining the mildly haemophilic phenotype observed in the proband from family 3. And in fact, *F8* sequencing revealed a hemizygous missense mutation that has been associated with mild haemophilia A. Hence, this is a clear example of how such duplications can confound the molecular diagnosis of haemophilia A via *F8* inversion analysis. As a result, in cases of a positive inversion analysis with weak clinical correlation, we recommend the use of FISH or array CGH to rule out an *int22h-1/int22h-2*-mediated Xq28 duplication.

In addition to the *F8* gene, which encodes coagulation factor VIII, the duplicated region contains nine other genes: *H2AFB*, *F8A*, *FUNDC2*, *MTCP1NB*, *MTCP1*, *BRCC3*, *VBP1*, *RAB39B* and *CLIC2* (<http://genome.ucsc.edu>). *H2AFB* genes encode members of the histone H2A family.⁴⁶ The *F8A* gene codes for the HAP40 protein, which co-purifies with the huntingtin protein, suggesting that HAP40 is likely to contribute to the function of normal huntingtin and is a candidate for involvement in the aberrant nuclear localisation of mutant huntingtin found in degenerating neurons in Huntington disease.⁴⁷ The *BRCC3* gene encodes a subunit of the BRCA1-BRCA2-containing complex, which is an E3 ubiquitin ligase. It is thought to be involved in the cellular response to ionising radiation and progression beyond the G2/M checkpoint. Therefore, it enhances cellular survival following DNA damage.⁴⁸ The *VBP1* gene encodes the Von Hippel–Lindau

binding protein-1. Von Hippel–Lindau syndrome is associated with mutations in the *VHL* gene, which encodes the tumour-suppressor VHL. It was found that VHL acts as a molecular chaperone responsible for carrying Von Hippel–Lindau binding protein-1 from perinuclear granules to the nucleus or cytoplasm.⁴⁹ Expression in mice during fetal stages was primarily limited to the central nervous system, retina and liver, whereas adult expression was more evenly distributed.⁵⁰ The *RAB39B* gene encodes a member of the Rab family of proteins, which are small GTPases that are involved in vesicular trafficking.⁵¹ *CLIC2* encodes the chloride intracellular channel 2 protein, which shares 60% sequence identity with the chloride intracellular channel 1 protein, a nuclear chloride channel.⁵² It is expressed in fetal liver and adult skeletal muscle and has been suggested to function as an intrinsic stabiliser of ryanodine receptors in skeletal muscles.⁵³ Nevertheless, none of the aforementioned genes have been shown to be associated with XLID, although increased dosage of one or more of those genes may affect brain development, resulting in cognitive impairment.

In conclusion, *int22h-1/int22h-2*-mediated Xq28 duplications may represent a novel XLID syndrome resulting in cognitive impairment, behavioural abnormalities, recurrent infections and characteristic facial features in males. Carrier females may exhibit a clinical phenotype with skewed XCI. Such duplications do not affect factor VIII levels, but can result in errors in the molecular diagnosis of haemophilia A. Alternatively, the reciprocal deletion in females may not have phenotypic consequences as it appears to result in completely skewed XCI. However, these deletions may be lethal for males in utero, resulting in recurrent spontaneous abortions in carrier females.

Competing interests None.

Ethics approval Institutional Review Board of Baylor College of Medicine.

Contributors Ayman El-Hattab gathered the clinical data and wrote the manuscript draft. Ping Fang and Weihong Jin performed the chromosome X-inactivation assays. James Gibson, Gayle Patel, Dorothy Grange and Linda Manwaring provided clinical information about the patients. Jeffrey Hughes, Ankita Patel and Pawel Stankiewicz reviewed the manuscript. Sau Wai Cheung is the senior author who identified the cases, designed the manuscript and reviewed it.

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REFERENCES

- Leonard H, Wen X. The epidemiology of mental retardation: challenges and opportunities in the new millennium. *Ment Retard Dev Disab Res Rev* 2002;**8**:117–34.
- Chiurazzi P, Schwartz CE, Gecz J, Neri G. XLMR genes: update 2007. *Eur J Hum Genet* 2008;**16**:422–34.
- Lisik MZ, Sieron AL. X-linked mental retardation. *Med Sci Monit* 2008;**14**:221–9.
- Ropers HH, Hamel BC. X-linked mental retardation. *Nat Rev Genet* 2005;**6**:46–57.
- Sharp AJ, Hansen S, Selzer RR, Cheng Z, Regan R, Hurst JA, Stewart H, Price SM, Blair E, Hennekam RC, Fitzpatrick CA, Segreaves R, Richmond TA, Guiver C, Albertson DG, Pinkel D, Eis PS, Schwartz S, Knight SJ, Eichler EE. Discovery of previously unidentified genomic disorders from the duplication architecture of the human genome. *Nat Genet* 2006;**38**:1038–42.
- Stankiewicz P, Beaudet AL. Use of array CGH in the evaluation of dysmorphology, malformations, developmental delay, and idiopathic mental retardation. *Curr Opin Gene Dev* 2007;**17**:182–92.
- Boone PM, Bacino CA, Shaw CA, Eng PA, Hixson PM, Pursley AN, Kang SH, Yang Y, Wisniewska J, Nowakowska BA, del Gaudio D, Xia Z, Simpson-Patel G, Immken LL, Gibson JB, Tsai AC, Bowers JA, Reimschuessel TE, Schaaf CP, Potocki L, Scaglia F, Gambin T, Sykulski M, Bartnik M, Derwinska K, Wisniewska-Kowalik B, Lalani SR, Probst FJ, Bi W, Beaudet AL, Patel A, Lupski JR, Cheung SW, Stankiewicz P. Detection of clinically relevant exonic copy-number changes by array CGH. *Hum Mutat* 2010;**31**:1326–42.
- Froyen G, Van Esch H, Bauters M, Hollanders K, Frints SG, Vermeesch JR, Devriendt K, Frys JP, Marynen P. Detection of genomic copy number changes in patients with idiopathic mental retardation by high-resolution X-array-CGH: important role for increased gene dosage of XLMR genes. *Hum Mut* 2007;**28**:1042–3.
- Van Esch H, Bauters M, Ignatius J, Jansen M, Raynaud M, Hollanders K, Lugtenberg D, Bienenvenu T, Jensen LR, Gecz J, Moraine C, Marynen P, Frys JP, Froyen G. Duplication of the *MECP2* region is a frequent cause of severe mental retardation and progressive neurological symptoms in males. *Am J Hum Genet* 2005;**77**:442–53.
- Friez MJ, Jones JR, Clarkson K, Lubs H, Abuelo D, Bier JA, Pai S, Simensen R, Williams C, Giampietro PF, Schwartz CE, Stevenson RE. Recurrent infections, hypotonia, and mental retardation caused by duplication of *MECP2* and adjacent region in Xq28. *Pediatrics* 2006;**118**:1687–95.
- Madrigal I, Rodríguez-Reuenga L, Armengol L, González E, Rodríguez B, Badenas C, Sánchez A, Martínez F, Guitart M, Fernández I, Arranz JA, Tejada M, Pérez-Jurado LA, Estivill X, Milà M. X-chromosome tiling path array detection of copy number variants in patients with chromosome X-linked mental retardation. *BMC Genomics* 2007;**8**:443.
- Breman AM, Ramocki MB, Kang SH, Williams M, Freedberg D, Patel A, Bader PI, Cheung SW. *MECP2* duplications in six patients with complex sex chromosome rearrangements. *Eur J Hum Genet* 2011;**19**:409–15.
- Froyen G, Bauters M, Boyle J, Van Esch H, Govaerts K, van Bokhoven H, Ropers HH, Moraine C, Chelly J, Frys JP, Marynen P, Gecz J, Turner G. Loss of *SLC38A5* and *FTSJ1* at Xp11.23 in three brothers with non-syndromic mental retardation due to a microdeletion in an unstable genomic region. *Hum Genet* 2007;**121**:539–47.
- Lugtenberg D, Yntema HG, Banning MJ, Oudakker AR, Firth HV, Willatt L, Raynaud M, Kleefstra T, Frys JP, Ropers HH, Chelly J, Moraine C, Gecz J, van Rieuwijk J, Nabuurs SB, de Vries BB, Hamel BC, de Brouwer AP, van Bokhoven H. *ZNF674*: a new Kruppel-associated box-containing zinc-finger gene involved in nonsyndromic X-linked mental retardation. *Am J Hum Genet* 2006;**78**:265–78.
- Van Esch H, Jansen A, Bauters M, Froyen G, Frys JP. Encephalopathy and bilateral cataract in a boy with an interstitial deletion of Xp22 comprising the *CDKL5* and *NHS* genes. *Am J Med Genet A* 2007;**143**:364–9.
- Froyen G, Corbett M, Vandewalle J, Jarvela I, Lawrence O, Meldrum C, Bauters M, Govaerts K, Vandeleur L, Van Esch H, Chelly J, Sanlaville D, van Bokhoven H, Ropers HH, Laumonnier F, Ranieri E, Schwartz CE, Abidi F, Tarpey PS, Futreal PA, Whibley A, Raymond FL, Stratton MR, Frys JP, Scott R, Peippo M, Sipponen M, Partington M, Mowat D, Field M, Hackett A, Marynen P, Turner G, Géc J. Submicroscopic duplications of the hydroxysteroid dehydrogenase *HSD17B10* and the E3 ubiquitin ligase *HUWE1* are associated with mental retardation. *Am J Hum Genet* 2008;**82**:432–43.
- Marshall CR, Noor A, Vincent JB, Lionel AC, Feuk L, Skaug J, Shago M, Moessner R, Pinto D, Ren Y, Thiruvahindrapuram B, Fiebig A, Schreiber S, Friedman J, Ketelaars CE, Vos YJ, Ficioglu C, Kirkpatrick S, Nicolson R, Sloman L, Summers A, Gibbons CA, Teebi A, Chitayat D, Weksberg R, Thompson A, Vardy C, Crosbie V, Luscombe S, Baatjes R, Zwaigenbaum L, Roberts W, Fernandez B, Szatmari P, Scherer SW. Structural variation of chromosomes in autism spectrum disorder. *Am J Hum Genet* 2008;**82**:477–88.
- Giorda R, Bonaglia MC, Beri S, Fichera M, Novara F, Magini P, Urquhart J, Sharkey FH, Zucca C, Grasso R, Marelli S, Castiglia L, Di Benedetto D, Musumeci SA, Vitello GA, Failla P, Reitano S, Avola E, Bisulli F, Tinuper P, Mastrangelo M, Focchi I, Spaccini L, Torniero C, Fontana E, Lynch SA, Clayton-Smith J, Black G, Jonveaux P, Leheup B, Seri M, Romano C, dalla Bernardina B, Zuffardi O. Complex segmental duplications mediate a recurrent dup(X)(p11.22-p11.23) associated with mental retardation, speech delay, and EEG anomalies in males and females. *Am J Hum Genet* 2009;**85**:394–400.
- Lugtenberg D, de Brouwer AP, Kleefstra T, Oudakker AR, Frints SG, Schrander-Stumpel CT, Frys JP, Jensen LR, Chelly J, Moraine C, Turner G, Veltman JA, Hamel BC, de Vries BB, van Bokhoven H, Yntema HG. Chromosomal copy number changes in patients with non-syndromic X linked mental retardation detected by array CGH. *J Med Genet* 2006;**43**:362–70.
- Jehee FS, Rosenberg C, Krepschi-Santos AC, Kok F, Knijnenburg J, Froyen G, Vianna-Morgante AM, Opitz JM, Passos-Bueno MR. An Xq22.3 duplication detected by comparative genomic hybridization microarray (array-CGH) defines a new locus (FGS5) for FG syndrome. *Am J Med Genet A* 2005;**139**:221–6.
- El-Hattab A, Bournat J, Eng P, Wu J, Walker B, Stankiewicz P, Cheung S, Brown C. Microduplication of Xp11.23p11.3 with effects on cognition, behavior, and craniofacial development. *Clin Genet*. Published Online First: 29 Jun 2010. doi:10.1111/j.1399-0004.2010.01496.x
- Cheng SF, Rauen KA, Pinkel D, Albertson DG, Cotter PD. Xq chromosome duplication in males: clinical, cytogenetic and array CGH characterization of a new case and review. *Am J Med Genet A* 2005;**135**:308–13.
- Sanlaville D, Schluth-Bolard C, Turleau C. Distal Xq duplication and functional Xq disomy. *Orphanet J Rare Dis* 2009;**4**:4.
- Vasquez AI, Rivera H, Bobadilla L, Crolla JA. A familial Xp+ chromosome, dup (Xq26.3qter). *J Med Genet* 1995;**32**:891–3.
- Goodman BK, Shaffer LG, Rutberg J, Leppert M, Harum K, Gagos S, Ray JH, Bialer MG, Zhou X, Pletcher BA, Shapiro SK, Geraghty MT. Inherited duplication Xq27-qter at Xp22.3 in severely affected males: molecular cytogenetic evaluation and clinical description in three unrelated families. *Am J Med Genet* 1998;**80**:377–84.
- Novelli A, Bernardini L, Salpietro DC, Briuglia S, Merlino MV, Mingarelli R, Dallapiccola B. Disomy of distal Xq in males: case report and overview. *Am J Med Genet* 2004;**128**:165–9.
- Sanlaville D, Prieur M, de Blois MC, Genevieve D, Lapierre JM, Ozilou C, Picq M, Gosset P, Morichon-Delvallez N, Munnich A, Cormier-Daire V, Baujat G, Romana S, Vekemans M, Turleau C. Functional disomy of the Xq28 chromosome region. *Eur J Hum Genet* 2005;**13**:579–85.
- El-Hattab AW, Smolarek TA, Walker ME, Schorry EK, Immken LL, Patel G, Abbott MA, Lanpher BC, Ou Z, Kang SH, Patel A, Scaglia F, Lupski JR, Cheung SW, Stankiewicz P. Redefined genomic architecture in 15q24 directed by patient deletion/duplication breakpoint mapping. *Hum Genet* 2009;**126**:589–602.
- Ou Z, Kang SH, Shaw CA, Carmack CE, White LD, Patel A, Beaudet AL, Cheung SW, Chinault AC. Bacterial artificial chromosome-emulsion oligonucleotide arrays for targeted clinical array-comparative genomic hybridization analyses. *Genet Med* 2008;**10**:278–89.
- Bernardini L, Sinibaldi L, Ceccarini C, Novelli A, Dalla Piccola B. Reproductive history of a healthy woman with mosaic duplication of chromosome 4p. *Prenat Diagn* 2005;**25**:283–5.
- Allen RC, Zoghbi HY, Moseley AB, Rosenblatt HM, Belmont JW. Methylation of *HpaII* and *HhaI* sites near the polymorphic CAG repeat in the human androgen-receptor gene correlates with X chromosome inactivation. *Am J Hum Genet* 1992;**51**:1229–39.
- Carrel L, Willard HF. An assay for X inactivation based on differential methylation at the fragile X locus, FMR1. *Am J Med Genet* 1996;**64**:27–30.
- Sharp A, Robinson D, Jacobs P. Age- and tissue-specific variation of X chromosome inactivation ratios in normal women. *Hum Genet* 2000;**107**:343–9.
- Liu Q, Nozari G, Sommer SS. Single-tube polymerase chain reaction for rapid diagnosis of the inversion hotspot of mutation in hemophilia A. *Blood* 1998;**92**:1458–9.
- Liu M, Murphy ME, Thompson AR. A domain mutations in 65 haemophilia A families and molecular modelling of dysfunctional factor VIII proteins. *Br J Haematol* 1998;**103**:1051–60.
- De Brasi CD, Bowen DJ. Molecular characteristics of the intron 22 homologs of the coagulation factor VIII gene: an update. *J Thromb Haemost* 2008;**6**:1822–4.
- Lakich D, Kazanian HH Jr, Antonarakis SE, Gitschier J. Inversions disrupting the factor VIII gene are a common cause of severe haemophilia A. *Nat Genet* 1993;**5**:236–41.
- Naylor J, Brinke A, Hassock S, Green PM, Giannelli F. Characteristic mRNA abnormality found in half the patients with severe haemophilia A is due to large inversions. *Hum Mol Genet* 1993;**2**:1773–8.
- Ross MT, Grafham DV, Coffey AJ, Scherer S, McLay K, Muzny D, Platzer M, Howell GR, Burrows C, Bird CP, Frankish A, Lovell FL, Howe KL, Ashurst JL, Fulton RS, Sudbrak R, Wen G, Jones MC, Hurler ME, Andrews TD, Scott CE, Searle S, Ramser J, Whittaker A, Deadman R, Carter NP, Hunt SE, Chen R, Cree A, Gunaratne P, Havlak P, Hodgson A, Metzker ML, Richards S, Scott G, Steffen D, Sodergren E, Wheeler DA, Worley KC, Ainscough R, Ambrose KD, Ansari-Lari MA, Aradhya S, Ashwell RI, Babbage AK, Bagguley CL, Ballabio A, Banerjee R, Barker GE, Barlow KF, Barrett IP, Bates KN, Beare DM, Beasley H, Beasley O, Beck A, Bethel G, Blechschmidt K, Brady N, Bray-Allen S, Bridgeman AM, Brown AJ, Brown MJ, Bonnin D, Bruford EA, Buhay C, Burch P, Burford D, Burgess J, Burrill W, Burton J,

- Bye JM, Carder C, Carrel L, Chako J, Chapman JC, Chavez D, Chen E, Chen G, Chen Y, Chen Z, Chinault C, Ciccocioppa A, Clark SY, Clarke G, Clee CM, Clegg S, Clerc-Blankenburg K, Clifford K, Cobley V, Cole CG, Conquer JS, Corby N, Connor RE, David R, Davies J, Davis C, Davis J, Delgado O, Deshazo D, Dharni P, Ding Y, Dinh H, Dodsworth S, Draper H, Dugan-Rocha S, Dunham A, Dunn M, Durbin KJ, Dutta I, Eades T, Ellwood M, Emery-Cohen A, Errington H, Evans KL, Faulkner L, Francis F, Frankland J, Fraser AE, Galgoczy P, Gilbert J, Gill R, Glöckner G, Gregory SG, Gribble S, Griffiths C, Grocock R, Gu Y, Gwilliam R, Hamilton C, Hart EA, Hawes A, Heath PD, Heitmann K, Hennig S, Hernandez J, Hinzmann B, Ho S, Hoffs M, Howden PJ, Huckle EJ, Hume J, Hunt PJ, Hunt AR, Isherwood J, Jacob L, Johnson D, Jones S, de Jong PJ, Joseph SS, Keenan S, Kelly S, Kershaw JK, Khan Z, Kioschis P, Klages S, Knights AJ, Kosiura A, Kovar-Smith C, Laird GK, Langford C, Lawlor S, Leversha M, Lewis L, Liu W, Lloyd C, Lloyd DM, Loulseged H, Loveland JE, Lovell JD, Lozado R, Lu J, Lyne R, Ma J, Maheshwari M, Matthews LH, McDowall J, McLaren S, McMurray A, Meidl P, Meitinger T, Milne S, Miner G, Mistry SL, Morgan M, Morris S, Müller I, Mullikin JC, Nguyen N, Nordsiek G, Nyakatura G, O'Dell CN, Okwuonu G, Palmer S, Pandian R, Parker D, Parrish J, Pasternak S, Patel D, Pearce AV, Pearson DM, Pelan SE, Perez L, Porter KM, Ramsey Y, Reichwald K, Rhodes S, Ridler KA, Schlessinger D, Schueler MG, Sehra HK, Shaw-Smith C, Shen H, Sheridan EM, Shownkeen R, Skuce CD, Smith ML, Sothoran EC, Steingrubner HE, Steward CA, Storey R, Swann RM, Swarbrick D, Tabor PE, Taudien S, Taylor T, Teague B, Thomas K, Thorpe A, Timms K, Tracey A, Trevanion S, Tromans AC, d'Urso M, Verdusco D, Villasana D, Waldron L, Wall M, Wang Q, Warren J, Warry GL, Wei X, West A, Whitehead SL, Whiteley MN, Wilkinson JE, Willey DL, Williams G, Williams L, Williamson A, Williamson H, Wilming L, Woodmansey RL, Wray PW, Yen J, Zhang J, Zhou J, Zoghbi H, Zorilla S, Buck D, Reinhardt R, Poustka A, Rosenthal A, Lehrach H, Meindl A, Minx PJ, Hillier LW, Willard HF, Wilson RK, Waterston RH, Rice CM, Vaudin M, Coulson A, Nelson DL, Weinstock G, Sulston JE, Durbin R, Hubbard T, Gibbs RA, Beck S, Rogers J, Bentley DR. The DNA sequence of the human X chromosome. *Nature* 2005;**434**:325–37.
40. **Bagnall RD**, Giannelli F, Green PM. Polymorphism and hemophilia A causing inversions in distal Xq28: a complex picture. *J Thromb Haemost* 2005;**3**:2598–9.
 41. **Bagnall RD**, Giannelli F, Green PM. Int22h-related inversions causing hemophilia A: a novel insight into their origin and a new more discriminant PCR test for their detection. *J Thromb Haemost* 2006;**4**:591–8.
 42. **Cullen CR**, Hubberman P, Kaslow DC, Migeon BR. Comparison of factor IX methylation on human active and inactive X chromosomes: implications for X inactivation and transcription of tissue-specific genes. *EMBO J* 1986;**5**:2223–9.
 43. **Pegoraro E**, Whitaker J, Mowery-Rushton P, Surti U, Lanasa M, Hoffman EP. Familial skewed X inactivation: a molecular trait associated with high spontaneous-abortion rate maps to Xq28. *Am J Hum Genet* 1997;**61**:160–70.
 44. **Freije D**, Schlessinger D. A 1.6-Mb contig of yeast artificial chromosomes around the human factor VIII gene reveals three regions homologous to probes for the DXS115 locus and two for the DXSY64 locus. *Am J Hum Genet* 1992;**51**:66–80.
 45. **Green P**, Hill M, Bowen DJ. the UK Haemophilia Centre Doctors' Organisation Haemophilia Genetics Network. Duplications involving *int22h-1* of the factor VIII gene: a cause for concern in genetic testing for hemophilia A? *J Thromb Haemost* 2007;**5**:2155–6.
 46. **Chadwick BP**, Willard HF. A novel chromatin protein, distantly related to histone H2A, is largely excluded from the inactive X chromosome. *J Cell Biol* 2001;**152**:375–84.
 47. **Peters MF**, Ross CA. Isolation of a 40-kDa huntingtin-associated protein. *J Biol Chem* 2001;**276**:3188–94.
 48. **Dong Y**, Hakimi MA, Chen X, Kumaraswamy E, Cooch NS, Godwin AK, Shiekhattar R. Regulation of BRCC, a holoenzyme complex containing *BRCA1* and *BRCA2*, by a signalosome-like subunit and its role in DNA repair. *Molec Cell* 2003;**12**:1087–99.
 49. **Tsuchiya H**, Iseda T, Hino O. Identification of a novel protein (VBP-1) binding to the von Hippel–Lindau (VHL) tumor suppressor gene product. *Cancer Res* 1996;**56**:2881–5.
 50. **Hemberger M**, Himmelbauer H, Neumann HPH, Plate KH, Schwarzkopf G, Fundele R. Expression of the von Hippel–Lindau-binding protein-1 (Vbp1) in fetal and adult mouse tissues. *Hum Molec Genet* 1999;**8**:229–36.
 51. **Cheng H**, Ma Y, Ni X, Jiang M, Guo L, Ying K, Xie Y, Mao Y. Isolation and characterization of a human novel RAB (RAB39B) gene. *Cytogenet Genome Res* 2002;**97**:72–5.
 52. **Heiss NS**, Poustka A. Genomic structure of a novel chloride channel gene, CLIC2, in Xq28. *Genomics* 1997;**45**:224–8.
 53. **Meng X**, Wang G, Viero C, Wang Q, Mi W, Su XD, Wagenknecht T, Williams AJ, Liu Z, Yin CC. CLIC2-RyR1 interaction and structural characterization by cryo-electron microscopy. *J Mol Biol* 2009;**387**:320–34.